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(54) **Artificial dura mater having cell adhesiveness and a process for producing the same**
Künstliche Dura Mater mit Zellhaftung und Prozess zur Produktion
Dure mère artificielle ayant une certaine adhesion cellulaire et son processus de fabrication

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(56) References cited:
DE-A- 19 904 913 US-A- 5 133 757
US-A- 5 308 704 US-A- 5 906 824
US-A- 6 051 751

• DATABASE MEDLINE [Online] August 1993
(1993-08) YAMAGATA S ET AL: "Clinical
experience with expanded
polytetrafluoroethylene sheet used as an
artificial dura mater." Database accession no.
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Description**TECHNICAL FIELD**

[0001] The present invention relates to a material having adhesiveness to bone and/or fascia which is useful as a medical material such as an artificial dura mater, and a process for producing the same.

BACKGROUND ART

[0002] Among three layers of membranes (pia mater, arachnoid, and dura mater) for protecting cerebral parenchyma in a cranial bone, the dura mater is the hardest outermost layer and is also the inner periosteum of the cranial bone. In brain neurosurgery, excision of the dura mater is often unavoidable, and this causes a defect in the dura mater. Spontaneous contraction of the dura mater itself sometimes complicates primary suturing. Occlusion of wound while the dura mater remains open results in serious complications, for example, intracranial infection caused by leakage of spinal fluid, local neurological symptom caused by adhesion between cerebral parenchyma and bones or subcutaneous tissues, and a focus of epileptic seizure. Therefore, when a wound is occluded, precise suturing is required in order not to create a gap in the dura mater. When a defect appears in the dura mater, or primary suturing is difficult, some prostheses should be used to completely suture without any gap.

[0003] Selection of a prosthesis for use in filling a dura mater defect is a problem which has troubled neurosurgeons for a long time. At an early stage, artifacts were used, however, there were problems such as biocompatibility and usability. Thus, no artifact has been used for a long period. Autogenous fascia is the most extensively used prosthesis from the early stage up to the present, but it has some problems such as production of a fascia defect on the extracted region and easy adhesion to a brain. A human dried dura mater is a prosthesis for the dura mater prepared by treating the dura mater collected from a corpse with radiation, and has been the best prosthesis among existing ones. However, prions which may be a cause of Creutzfeldt Jakob disease, are potentially present in the dura mater, and infection of Creutzfeldt Jakob disease through a human dried dura mater has been reported. Therefore, use thereof was completely prohibited in 1998.

[0004] The only material usable as a prosthesis for the dura mater in addition to autogenous fascia is ePTFE (expanded polytetrafluoroethylene), which has been approved by Department of Health and Human Services. Because ePTFE is a polymeric material, it has no adhesiveness to organisms. This property is excellent in that ePTFE does not adhere to the brain. On the other hand, ePTFE is poor in contractility, and spinal fluid can leak from a pinhole, thereby requiring suturing with the use of special surgical suture. Since there is no adhesiveness

to organisms, spinal fluid is highly likely to leak from a gap between suture surfaces. In addition, since there is no adhesiveness to peripheral tissue, it is highly likely to be no more than a skeletal material. Many attempts have been heretofore made on how to successfully use ePTFE. However, all of these attempts involved using ePTFE as a skeletal material and waiting until a coat of fibrous tissue was formed on the periphery.

SUMMARY OF THE INVENTION

[0005] The present inventors have heretofore conducted various experiments by modifying a surface layer of ePTFE, polylactic acid or Polyglactin by the ion implantation technique and changing the cell adhesiveness, and have examined utility values thereof. The present inventors have aimed at development of a novel artificial dura mater with one side having no cell adhesiveness and the other side having cell adhesiveness, by means of surface modification of the polymeric material by the ion implantation technique.

[0006] When a polymeric material is irradiated with an ion beam, the admitted ion occasionally loses energy by collision with a matrix atom and finally stops. Over the course thereof, bond cleavage or new bonding occurs. In general, when nuclear stopping power is governing, occurrence of defects such as radicals or change in conformation, density, and morphology, is observed. In contrast, when electron stopping power is governing, electronic excitation ensures, and cleavage or crosslinking of a branched-chain is thought to occur. Interatomic bonds of polymeric materials have a binding energy of about several eV, and the amount of energy per unit length to be applied to the matrix by ion beam irradiation is several tens of eV/Å. That is, it is the application of an energy flux ten times or larger than the binding energy. Change in physical property of polymeric materials by the irradiation of this energy flux has been observed. These phenomena vary depending on the type of ion beam, acceleration energy, amount of irradiation, and type of matrix. When polymeric materials are irradiated with an ion beam, beam current should not be excessively raised because polymeric materials have a low tolerance for heat. Also, since polymeric materials are electrical insulants, attention should be paid to the possibility that an accurate applied amount may not be measured by current measurement.

[0007] The present inventors have heretofore conducted surface modification by irradiating ePTFE, polylactic acid and Polyglactin materials with ion species such as helium (He) and neon (Ne), and have repeated basic experiments relating to cell adhesiveness by using a cultured cell system. As a result, it has been clarified that cell adhesiveness can be imparted by irradiation of Neon beam at an acceleration energy of 150 keV and at an ion beam current of 0.5 $\mu\text{A}/\text{cm}^2$ or below to ePTFE, the application of which to human has been approved. Also, it has been clarified that cell adhesiveness varies de-

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pending on the amount of ion irradiation, and sufficient adhesiveness cannot be attained with an excessively large or small amount of irradiation. In hitherto experiments, cell adhesiveness was observed on the Ne-ion irradiated ePTFE surface at a dose of 1×10^{14} ions/cm², and adhesiveness was the highest at a dose of around 1×10^{15} ions/cm² while cell adhesiveness was lowered at a dose of 1×10^{17} ions/cm². Amorphous carbonization by ion beam irradiation was heretofore considered to enhance adhesiveness. However, cell adhesion was observed even at the dose of 1×10^{14} ions/cm². The amount added at this dose is insignificant based on a theoretical calculation, and irradiation damage is at a level that cannot be detected by analysis such as Fourier transform infrared spectroscopy or Raman spectroscopy. Cells detect such a subtle change, and adhere. How cells detect this subtle change, and the mechanism of adhesion are not yet elucidated, and study thereof is being conducted using various analytical instruments.

[0008] ePTFE, polylactic acid and Polyglactin materials to which an ion beam has been applied show improvement in cell adhesiveness as well as adhesion strength. Cells allow substances similar to octopus legs referred to as "desmosomes" to invade onto a surface of the artificial material, thereby immobilizing themselves. It is predicted that adhesion strength on the ion beam irradiated surface is probably improved by increasing desmosomes. It has been confirmed that ion beam irradiated polymeric materials has a smooth surface, so long as the region was observed under an atomic force microscope, and it is not because the surface is roughened due to ion beam irradiation and the adhesion area with cells enlarged, as initially predicted. It has been confirmed in a basic experiment using cultured cells that desmosomes are increased by selecting surface modification conditions.

[0009] By determining an optimal dose of ion beam to ePTFE, which is applicable to organisms, based on the data collected by hitherto basic experiments, clinical application can presumably be made possible. At an early stage, ePTFE, the surface of which has been treated at the optimal dose, only played a role as an artificial dura mater for filling a defected portion on the dura mater in cranial neurosurgery. However, as considering the fact that the human dried dura mater has been used as a prosthesis used in skull base formation in skull base surgery, as a prosthesis in the surgical operation of spinal disease, and also as a prosthesis in other surgery on an abdominal or thoracic organ or musculoskeleton, the present inventors have come to consider the existence of a more extensive scope of application. In any site, if tissue adhesion is not desired on one side while tissue adhesion is desired on the other side, this Ne-ion beam-irradiated ePTFE can be widely used.

[0010] However, the above-mentioned data on cell adhesiveness of ePTFE is merely collected *in vitro*, i.e., in a cultured cell system. Thus, a degree of *in vivo* cell adhesiveness actually exhibited by Ne-ion beam-irradiated

ePTFE has not been elucidated. Therefore, it is not clear whether or not the optimal ion beam dose of 1×10^{15} ions/cm² that has been obtained in a cultured cell system is the optimal dose *in vivo*. There has been heretofore no report on types of morphological, histochemical conditions caused by cell adhesiveness that would occur *in vivo*, and in particular, on dynamics of desmosomes. Since a self-control system, that is, a tissue repair mechanism, is active *in vivo*, the occurrence of a situation is not hard to imagine which is unexplainable from a simple reaction such as in the cultured cell system. Also, there is a lot of unknown problems including the clinical significance of the reinforcement of cell adhesiveness that would actually occur *in vivo*, the problem of biocompatibility, the operability premising the surgical operation, and the occurrence of a reaction against foreign body.

[0011] Specifically, the object of the present invention is to elucidate the manner in which ePTFE adheres to cells together with morphological and histochemical changes over the period of time, by allowing ePTFE which has been irradiated with an ion beam under certain condition to closely adhere to bones and muscles, and also to examine whether or not this cell adhesiveness is enduring in the clinical application.

[0012] The present inventors have conducted concentrated studies in order to attain the above object, and as a result, they found that Ne-ion beam-irradiated ePTFE had cell adhesiveness even *in vivo*, thereby completing the present invention.

[0013] Thus, according to the present invention, there is provided a material having adhesiveness to bone and/or fascia, which is composed of a polymeric material comprising carbon as a constituent element out of the group ePTFE, polylactic acid and Polyglactin and at least a part of a surface of which is modified by ion bombardment.

[0014] The polymeric material comprising carbon as a constituent element are expanded polytetrafluoroethylene (ePTFE), polylactic acid, or Polyglactin; and the above-mentioned materials wherein modification by ion bombardment is performed by ion implantation in a dose ϕ range of $1 \times 10^{13} \leq \phi < 1 \times 10^{16}$ ions/cm².

[0015] According to another aspect of the present invention, there is provided a material for an artificial dura mater which is composed of the material having adhesiveness to bone and/or fascia as mentioned above.

[0016] According to further another aspect of the present invention, there is provided a process for producing a material having adhesiveness to bone and/or fascia wherein ion implantation is performed in a dose ϕ range of $1 \times 10^{13} \leq \phi < 1 \times 10^{16}$ ions/cm² on at least a part of a surface of a polymeric material comprising carbon as a constituent element out of the group ePTFE, polylactin acid and Polyglactin.

BRIEF DESCRIPTION OF DRAWINGS

[0017]

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Fig. 1 shows morphological changes in ePTFE on fascia wherein:

Fig. 1-1 : changes on muscle on the second week at a dose of 1×10^{14} ions/cm²; Adhesion is observed between a small number of histiocytes and fibroblasts on the irradiated surface, while no cell adhesion is observed at all on the unirradiated surface.

Fig. 1-2 : changes on muscle on the fourth week at a dose of 1×10^{14} ions/cm²; Strong adhesion of fibroblasts and desmosomes are observed on the irradiated surface, while no cell adhesion is observed at all on the unirradiated surface.

Fig. 1-3 : changes on muscle on the second week on an unirradiated surface; Accumulation of histiocytes is observed on the surface which is in contact with ePTFE, i.e., the outermost layer of the fibrous tissue, and

Fig. 1-4 shows changes on muscle on the fourth week on the unirradiated surface. A solid coating of fibrous tissue is formed so as to cover ePTFE. Histiocytes still remain in the portion in contact with ePTFE.

Fig. 2 shows morphological changes in ePTFE on fascia and bones wherein

Fig. 2-1 shows changes on muscle on the second week at a dose of 5×10^{14} ions/cm²; Strong adhesion between histiocytes and fibrous tissues on the surface of ePTFE, a small number of desmosomes, and penetration of liquid component are observed.

Fig. 2-2 shows changes on muscle on the fourth week at a dose of 5×10^{14} ions/cm²; Strong fibrous adhesion is observed on ePTFE and the number of desmosomes is also increased.

Fig. 2-3 shows changes on bone on the second week at a dose of 1×10^{14} ions/cm²; ePTFE is in contact with bones, however, fibroblasts are present therebetween; and

Fig. 2-4 shows changes on bone on the second week at a dose of 5×10^{14} ions/cm²; Strong fibrous adhesion, and desmosomes are present on ePTFE. Penetration of liquid component is also significant.

Fig. 3 shows morphological changes in ePTFE on fascia and bones wherein

Fig. 3-1 shows changes on muscle on the fourth week at a dose of 1×10^{15} ions/cm²; Significant level of penetration of liquid component and thick cell adhesion are observed. Expression of phagocytes is also observed.

Fig. 3-2 shows changes on muscle on the fourth week at a dose of 1×10^{15} ions/cm²; There is

substantially no penetration of liquid component. Histiocytes were expressed on the surface of the muscular tissue, and solid cell adhesion is created.

Fig. 3-3 shows changes on bone on the second week at a dose of 1×10^{15} ions/cm²; Various cell components is adhered to ePTFE. Many desmosomes are observed, and penetration of liquid component is also significant, and

Fig. 3-4 shows changes on bone on the fourth week at a dose of 1×10^{14} ions/cm². Long desmosomes are observed on ePTFE, while penetration of liquid component is insignificant.

Fig. 4 shows morphological changes in ePTFE on bones wherein

Fig. 4-1 shows changes on bone on the fourth week at a dose of 5×10^{14} ions/cm²; Liquid component is penetrated into a considerably deep level. Long legs of desmosomes are observed along therewith.

Fig. 4-2 shows changes on bone on the fourth week at a dose of 1×10^{15} ions/cm²; Deep, wide, and accumulated desmosomes. Penetration of liquid component is significant.

Fig. 4-3 shows changes on bone on the second week at a dose of 1×10^{15} ions/cm²; The bone tissue is present in contact with ePTFE, and a small number of histiocytes and fibroblasts are present therebetween, and

Fig. 4-4 shows changes on bone on the fourth week at a dose of 1×10^{16} ions/cm². There are substantially no histiocytes or fibroblasts, and the bone tissue is in direct contact with ePTFE. Liquid component is not substantially penetrated, and existence of desmosomes is not clear.

EMBODIMENT FOR CARRYING OUT THE INVENTION

[0018] The embodiment for carrying out the present invention will be described below in detail.

[0019] The polymeric materials used in the present invention are expanded polytetrafluoroethylene (ePTFE), polylactic acid and Polyglactin. Expanded polytetrafluoroethylene (ePTFE) is particularly preferred.

[0020] At least a part of the surface of the polymeric material having adhesiveness to bones and/or muscles of the present invention is modified by ion bombardment. Examples of ionic species to be implanted include He⁺, C⁺, N⁺, Ne⁺, Na⁺, N₂⁺, O₂⁺, Ar⁺, and Kr⁺, although the ionic species are not particularly limited to these unless the ionic species inhibit cell growth by elution.

[0021] The dose ϕ is preferably in the range of $1 \times 10^{13} \leq \phi < 1 \times 10^{16}$ ions/cm². When the dose is lower than 10^{13} ions/cm², a significant improvement effect of cell adhesiveness is lowered. When the dose is higher than 10^{16} ions/cm², polymeric materials are likely to be dam-

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aged. Thus, neither thereof is preferred. The dose ϕ is more preferably in the range of $1 \times 10^{13} \leq \phi < 1 \times 10^{15}$ ions/cm².

[0022] Regarding the ion accelerating energy, its rise and fall may be causative of differences in the energy transfer mechanism. In practice, the energy level can be set in the range of about several tens to several hundreds keV, preferably about 50 to 150 keV.

[0023] The beam current density is preferably set in a range not exceeding approximately 0.5 $\mu\text{A}/\text{cm}^2$. This is because of a fear that an excessive beam current density excessively raises temperature of a target polymeric material, thereby deteriorating the polymeric material itself and lowering cell adhesiveness.

[0024] According to the present invention, means for imparting ion bombardment include ion implantation. In ion implantation, the reaction itself is limited to interaction between the ion beam and the material to be implanted (a target material). In addition, ions can be embedded at a desired depth from the surface by selecting ion implantation energy, and the controllability is excellent. This is a feature being absent in the plasma treatment. The implanted ions have mechanical differences, specifically, electron stopping power functions on ions having relatively low mass at the initial stage of diffusion, while nuclear stopping power functions on ions having relatively high mass from the outset. However, the implanted ions allow polymeric materials to be heated by lattice vibration (a state of thermal non-equilibrium), giving rise to melting, amorphization, and the like.

[0025] The present invention will be described in more detail with reference to the following examples although the present invention is not limited by these examples.

EXAMPLES

(A) Summary of the experiment

(1) Correlation between organism adhesiveness and ion dose of ePTFE having the surface modified by Ne-ion beam-irradiation

[0026] The results of hitherto experiments using a cultured cell system revealed that cell adhesiveness was attained at a dose of 1×10^{14} ions/cm², the maximal cell adhesiveness was attained at a dose of 1×10^{15} ions/cm², and cell adhesiveness was significantly lost at a dose of 1×10^{16} ions/cm². In the *in vivo* implantation experiment, ePTFE at a dose of 1×10^{14} ions/cm², ePTFE at a dose of 5×10^{14} ions/cm², and ePTFE at a dose of 1×10^{15} ions/cm² were used to observe and study variation of cell adhesiveness depending on the dose.

(2) Changes with time in organism adhesion of ePTFE having the surface modified by Ne beam irradiation

[0027] Cells are always acting dynamically *in vivo*, and an organism controlling system referred to as a mecha-

nism of wound healing is operating. Therefore, even on the surface of the ion beam-irradiated surface-modified ePTFE, which has been implanted in the organism, cell adhesion would proceed with time. Observation for an ultra-short-term to an ultra-long-term is ideal, but it is difficult. Thus, observation of a specimen was concentrated and examined on the second week in which the primary adhesion of wound is approximately completed and the forth week in which a mechanism of repairing in tissue is presumably enhanced. Also in this case, since variation depending on the dose arises additionally, ePTFE at a dose of 1×10^{14} ions/cm², ePTFE at a dose of 5×10^{14} ions/cm², and ePTFE at a dose of 1×10^{15} ions/cm², were respectively observed with time.

(3) Correlation of adhesiveness between ePTFE having the surface modified by Ne beam irradiation and different tissues in organisms

[0028] One object of the present invention is to develop an artificial dura mater which, on filling a defect in the dura mater, has no cell adhesiveness on the surface contacting with the brain, but has cell adhesiveness on the surface contacting with cranial bone. However, when a human dried dura mater was used in the past, the human dried dura mater had been used in orthopedics, urological section, surgery and the like in addition to as a filling for the dura mater. Thus, in order to confirm adhesiveness in other organs which is feasible in the same experiment system, an experiment for adhesion to rabbit muscles was also carried out.

(B) Method for experiment

(1) Sample

[0029] Three types of material were provided for use by irradiating 20 mm-square ePTFE having a thickness of 0.3 mm with Ne-ion at an acceleration energy of 150 keV, at the dose of 1×10^{14} ions/cm², 5×10^{14} ions/cm², and 1×10^{15} ions/cm², respectively, at the ion beam current of 0.5 $\mu\text{A}/\text{cm}^2$ or below, using a 200 kV ion implantation apparatus (The Institute of Physical and Chemical Research). These three types of dose were selected based on the result of the basic experiment using a cultured cell system in which, cell adhesiveness to ePTFE was developed at the ion dose of 1×10^{14} ions/cm², the adhesion effect became maximum at 1×10^{15} ions/cm², and most of the adhesion effect was lost at 1×10^{16} ions/cm². In the actual experiment, materials to be irradiated were cut to a size depending on the size of embedded portion in such a way that the material was unlikely to be influenced by ambient forces.

(2) Animal test

[0030] 10 male Japanese white rabbits weighing 2.5 to 3.0 kg were used in the experiment. Under general

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anesthesia with isoflurane, 2% xylocaine was locally injected, and the scalp was incised along the coronal suture to expose the cranial bone. With the periosteum on the surface of the cranial bone being completely removed, a sample was placed on the bone side with the irradiated surface facing the bone side. Since there is no muscular tissue on the bone, the possibility of the sample moving is very low and thus, immobilization was not particularly carried out. The reason why the periosteum was removed is that the artificial dura mater filled in the usual operation is in direct contact with bones having no periosteum since the dura mater is responsible for a part of the periosteum in the cranial bone.

[0031] Subsequently, 2% xylocaine was placed on the dorsal paramedian and skin was vertically incised under local anesthesia, and subcutaneous tissue was bluntly ablated to expose the back group of muscles. Fascia in the back group of muscles were carefully ablated so as not to damage muscles, a sample was placed so as to apply the irradiated surface on the muscular coat, and four corners were lightly sutured and immobilized on fascia. It is because, in portions in which muscles are present, a sample is highly likely to move accompanying the movement of the muscle.

(3) Observation

[0032] Two weeks and four weeks after the sample was embedded and implanted, rabbits were sacrificed using Nembutal, and peripheral tissue was respectively extracted as a mass, and immobilized with 10% buffer formalin. A peripheral tissue was extracted in order to avoid separation between ePTFE and a tissue, because tissue does not adhere to ePTFE on the surface which is not treated with the ion beam. The sample placed on the muscle was embedded in paraffin, while the sample placed on the cranial bone was embedded in paraffin after decalcification of cranial bone. The samples were stained by hematoxylin-eosin and by Masson's trichrome, and were observed under a microscope.

(C) Result

(1) Morphological change in ePTFE placed on fascia

[0033] As previously reported, cell adhesion was not observed at all on the untreated ePTFE surface on the second and fourth weeks (Figs. 1-1 and 1-2 of Fig. 1). However, this does not indicate that there is no reaction at all in peripheral tissues. It seems that a smooth-surfaced fibrous tissue covers ePTFE interposing a narrow gap with ePTFE, on the second week, however, accumulation of histiocytes was observed on the surface of the fibrous tissue contacting ePTFE (Fig. 1-3 of Fig. 1). The present experiment could not clarify whether this accumulation developed as reactivity against ePTFE or a simple reaction of impaired tissue stump. On the fourth week, a fibrous component had increased in a histiocytes

accumulated site, fibroblasts on the periphery had thickened, and a solid coating had formed (Fig. 1-4 of Fig. 1).

[0034] In contrast, adhesion of cells which were considered to be fibroblasts, was observed on the surface of a coating two weeks after implantation on the Ne-ion beam treated ePTFE surface, and cellular infiltration of histiocytes, lymphocyte or the like was observed inside the fibrous coating provided on the periphery (Fig. 2-1 of Fig. 2).

[0035] Penetration of liquid component into ePTFE was observed on the side of a treated surface. Such penetration of liquid component was not observed on the untreated surface. Penetrated liquid component presumably contains proteins from its stainability, although types of component actually penetrated are unclear and should be examined. Desmosomes, directly associated with adhesion strength between ePTFE and the adhered surface, were also observed, and exhibited a tendency in which desmosomes increased on the fourth week compared with the second week (Fig. 2-2 of Fig. 2).

[0036] The accessibility between a fibroblast and an ion beam treated ePTFE surface was very low at 1×10^{14} ions/cm², penetration of liquid component was little, and expression of desmosomes was also low. As the dose increased from 5×10^{14} ions/cm² to 1×10^{15} ions/cm², penetration of liquid component increased, and desmosomes tended to increase (Fig. 3-1 of Fig. 3). However, even though penetration of liquid component was not observed in some portions, there was a portion in which desmosomes were observed. Thus, penetration of liquid component and expression of desmosomes were not always in a parallel correlation (Fig. 3-2 of Fig. 3).

[0037] A similar finding was obtained from a specimen extracted on the fourth week. ePTFE irradiated at a dose of 1×10^{14} ions/cm², exhibited adhesiveness equivalent to that at the second week, and the reactive fibrous component on the periphery had thickened. On ePTFE surfaces irradiated at 5×10^{14} ions/cm² and at 1×10^{15} ions/cm², cell adhesion had advanced compared with that at the second week, and desmosomes also tended to increase. Regarding the length of desmosomes, there was a portion exceeding 4000Å, which was the offset of Ne-ion beam into ePTFE at 150 keV.

(2) Morphological change in ePTFE placed on the cranial bone

[0038] There was no difference in reaction on the untreated ePTFE surface from the case where ePTFE was placed on the fascia. Regarding the specimen extracted on the second week, cell adhesiveness was observed at 1×10^{14} ions/cm² on the ion beam treated ePTFE surface that is in contact with bone, reaction of histiocytes and penetration of liquid component were observed, and a small number of desmosomes was also observed (Fig. 2-3 of Fig. 2). Compared with the case where ePTFE was placed on fascia, cell adhesion was impressively strong although it could not be objectively proved. As the dose

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increase from 5×10^{14} ions/cm² to 1×10^{15} ions/cm², contact of fibroblasts increased, and the reaction of histiocytes, penetration of liquid component and desmosomes tended to increase (Fig. 2-4 of Fig. 2, Fig. 3-3 of Fig. 3)

[0039] In the specimen extracted on the fourth week, although the group irradiated at 1×10^{14} ions/cm² exhibited a tendency to increase the thickness of fibroblasts in contact, penetration of liquid component did not significantly increase (Fig. 3-4 of Fig. 3). Regarding groups irradiated at 5×10^{14} ions/cm² and 1×10^{15} ions/cm², respectively, contact of fibroblasts on the ePTFE surface increased, and ePTFE desmosomes increased as with muscle surface, and accumulation of desmosomes was observed on a part (Figs. 4-1 and 4-2 of Fig. 4). Penetration of liquid component was also considered to increase. On both the second and the fourth weeks, expression of histiocytes and fibroblasts was low and an image was observed in which an osteoblast-like tissue was in direct contact with ePTFE (Figs. 4-3 and 4-4 of Fig. 4). However, it was not observed that this cell expressed desmosomes in ePTFE, and it could not be confirmed whether or not this was only simple contact.

[0040] The possibility was suggested that fibroblasts being in contact with an ion beam irradiated surface, produced desmosomes and invaded into ePTFE, and the distance exceeded the offset of the ion beam. This suggests that even if ePTFE does not have cell adhesiveness, there is a possibility for fibroblasts and the like to invade into ePTFE once the ePTFE grasps a chance. This also suggests a possibility of attaining a strong adhesiveness between ePTFE and cells advancing on the periphery.

(D) Conclusion and Discussion

[0041] A series of the present experiments clarified the following.

(1) Ne-ion beam-irradiated ePTFE has cell adhesiveness *in vivo*.

[0042] Experiments using a cultured cell system have demonstrated that Ne-ion beam-irradiated ePTFE has cell adhesiveness. Since a more complicated mechanism of wound healing actually exists *in vivo*, it was not clear how an organism reacted against Ne-ion beam-irradiated ePTFE. The present experiment have demonstrated that, in many cases, histiocytes, and in part, fibroblasts adhered to the treated surface on the second week regardless the amount of ion beam irradiated. In a small number of cases, it was observed that bone and muscle were in direct contact with bone and muscle without intermediating histiocytes or fibroblasts. Therefore, there are two possible cases in adhesion between a tissue and Ne-ion beam-irradiated ePTFE, i.e., a case where a cell adhesion factor, mainly consisting of histiocytes, plays a main role and a case where a peripheral

tissue directly adheres.

(2) Cell adhesiveness of Ne-ion beam-irradiated ePTFE *in vivo* depends on the amount of ion beam irradiated.

[0043] It is reported that there is a clear correlation between cell adhesiveness to Ne-ion beam-irradiated ePTFE and the amount of ion beam irradiated in the cultured cell system. However, it was not clear whether or not this condition also holds true *in vivo*. A series of the present experiments demonstrated that, as with the cultured cell system, the ion beam irradiated ePTFE had cell adhesiveness *in vivo* and strongly adhered through desmosomes. *In vivo*, however, appearance of cells such as histiocytes or phagocyte, which are not observed in the cultured cell system, was suggested to have strong correlation with adhesiveness. Cell adhesiveness of Ne-ion beam-irradiated ePTFE was clearly increased as the dose increased from 1×10^{14} , 5×10^{14} to 1×10^{15} , based on the condition of desmosome appearance and the like, as with the cultured cell system.

(3) Desmosomes play a key role in cell adhesion to Ne-ion beam-irradiated ePTFE.

[0044] Observation of the adhered surface between Ne-ion beam-irradiated ePTFE and a peripheral tissue in the present experiment demonstrated four types of adhesion conditions: a) histiocytes adhered on the surface; b) fibroblasts adhered; c) bone and muscle existing on the periphery adhered; and d) desmosomes appeared from histiocytes and fibroblasts and adhered ePTFE as if putting legs into ePTFE. Although peel strength was not measured in each condition, strong adhesiveness cannot be expected from mere existence of histiocytes or fibroblasts on ePTFE. This is because a similar finding can be obtained even on the surface that is not treated with the ion beam. Since adhesion to ePTFE is not observed on the unirradiated surface, if histiocytes and fibroblasts seem to be merely resting on Ne-ion beam-irradiated ePTFE, small desmosomes are highly likely to be present.

(4) Liquid component is penetrated into Ne-ion beam-irradiated ePTFE.

[0045] Based on the present experiment, from the manner of coloring, it was found that a liquid component presumably containing proteins had penetrated into Ne-ion beam-irradiated ePTFE. There were sites where the liquid component had penetrated from the surface over a wide range, and sites where small fistula were present and the liquid component had penetrated from the surface to inside while exhibiting a large spread.

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(5) Reaction on Ne-ion beam-irradiated ePTFE adhered surface is not always in parallel to the amount of ion beam irradiated, tissue in contact, and reaction time.

[0046] As a whole, a reaction on the adhered surface is enhanced at the optimal amount of irradiation, reinforced with the elapse of time, and develops a different reaction manner depending on the tissue in contact. However, when examining the experiment result in detail, conditions of histiocyte appearance on the adhered surface, conditions of fibroblast appearance, conditions of desmosome appearance and the like are not always in parallel to the amount of ion beam irradiation, the tissue in contact, reaction time and the like.

[0047] The result of the present *in vivo* implantation experiment using rabbits demonstrated sufficient cell adhesiveness, that is, Ne-ion beam-irradiated ePTFE exhibited a maximal effect at the dose of 1×10^{15} ions/cm². Also, substantially equivalent adhesiveness was exhibited to bone and muscle. These results indicate that Ne-ion beam-irradiated ePTFE is applicable in various clinical fields as an artificial dura mater as a prosthesis for dura mater, as well as a prosthesis for organism membrane having a bilateral character of adhesiveness and nonadhesiveness.

INDUSTRIAL APPLICABILITY

[0048] The present invention provides an artificial dura mater having cell adhesiveness and a process for producing the same. Specifically, the present invention realizes formation of the artificial dura mater having a function closer to that of an organism by imparting cell adhesiveness on the surface which is in contact with a cranial bone side. The present invention is further applicable as a prosthesis in surgery of spinal diseases and as a prosthesis in surgery on thoracic organs and abdominal organs or musculoskeleton in addition to the application thereof to the artificial dura mater.

Claims

1. A material having adhesiveness to bone and/or fascia, which is composed of a polymeric material comprising carbon as a constituent element, and at least a part of a surface of which is modified by ion bombardment, wherein the polymeric material comprising carbon as a constituent element is selected from the group expanded polytetrafluoroethylene (ePTFE), polylactic acid, or Polyglactin.
2. The material having adhesiveness to bone and/or fascia according to claim 1, wherein modification by ion bombardment is performed by ion implantation in a dose Φ range of $1 \times 10^{13} \leq \Phi < 1 \times 10^{16}$ ions/cm².
3. A material for an artificial dura mater which is com-

posed of the material having adhesiveness to bone and/or fascia according to claim 1.

4. A process for producing a material having adhesiveness to bone and/or fascia wherein ion implantation is performed in a dose Φ range of $1 \times 10^{13} \leq \Phi < 1 \times 10^{16}$ ions/cm² on at least a part of a surface of a polymeric material comprising carbon as a constituent element, wherein the polymeric material comprising carbon as a constituent element is selected from the group expanded polytetrafluoroethylene (ePTFE), polylactic acid, or Polyglactin.

Patentansprüche

1. Material mit Haftfähigkeit an Knochen und/oder Faszie, das aus einem polymeren Material zusammengesetzt ist, das Kohlenstoff als Bestandteil umfasst, und wenigstens ein Teil einer Oberfläche desselben durch Ionenbeschuss modifiziert ist, wobei das Kohlenstoff als Bestandteil umfassende polymere Material ausgewählt ist aus der Gruppe expandiertes Polytetrafluorethylen (ePTFE), Polymilchsäure oder Polyglactin.
2. Material mit Haftfähigkeit an Knochen und/oder Faszie nach Anspruch 1, wobei die Modifizierung durch Ionenbeschuss mittels Ionenimplantation in einem Bereich der Dosis Φ von $1 \cdot 10^{13} \leq \Phi < 1 \cdot 10^{16}$ Ionen/cm² durchgeführt wird.
3. Material für eine künstliche Dura mater, das zusammengesetzt ist aus dem Material mit Haftfähigkeit an Knochen und/oder Faszie nach Anspruch 1.
4. Verfahren zur Herstellung eines Materials mit Haftfähigkeit an Knochen und/oder Faszie, wobei die Ionenimplantation in einem Bereich der Dosis Φ von $1 \cdot 10^{13} \leq \Phi < 1 \cdot 10^{16}$ Ionen/cm² auf wenigstens einem Teil einer Oberfläche eines polymeren Materials durchgeführt wird, das Kohlenstoff als Bestandteil umfasst, wobei das Kohlenstoff als Bestandteil umfassende polymere Material ausgewählt ist aus der Gruppe expandiertes Polytetrafluorethylen (ePTFE), Polymilchsäure oder Polyglactin.

Revendications

1. Matériau ayant une adhésivité à l'os et/ou au fascia, qui est composé d'un matériau polymère comprenant du carbone comme élément constitutif et dont au moins une partie de surface est modifiée par bombardement ionique, dans lequel le matériau polymère comprenant du carbone comme élément constitutif est choisi dans le groupe constitué par le polytétrafluoroéthylène expansé (ePTFE), l'acide poly-

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lactique ou la polyglactine.

2. Matériau ayant une adhésivité à l'os et/ou au fascia selon la revendication 1, dans lequel on effectue une modification par bombardement ionique par implantation d'ions dans une plage de doses ϕ de 1×10^{13} $\phi < 1 \times 10^{16}$ ions/cm². 5
3. Matériau pour dure-mère artificielle qui est composé du matériau ayant une adhésivité à l'os et/ou au fascia selon la revendication 1. 10
4. Procédé pour produire un matériau ayant une adhésivité à l'os et/ou au fascia, dans lequel on effectue l'implantation d'ions dans une plage de doses ϕ de 1×10^{13} $\phi < 1 \times 10^{16}$ ions/cm² sur au moins une partie de la surface d'un matériau polymère comprenant du carbone comme élément constitutif, dans lequel le matériau polymère comprenant du carbone comme élément constitutif est choisi dans le groupe constitué par le polytétrafluoroéthylène expansé (ePTFE), l'acide polylactique ou la polyglactine. 15 20

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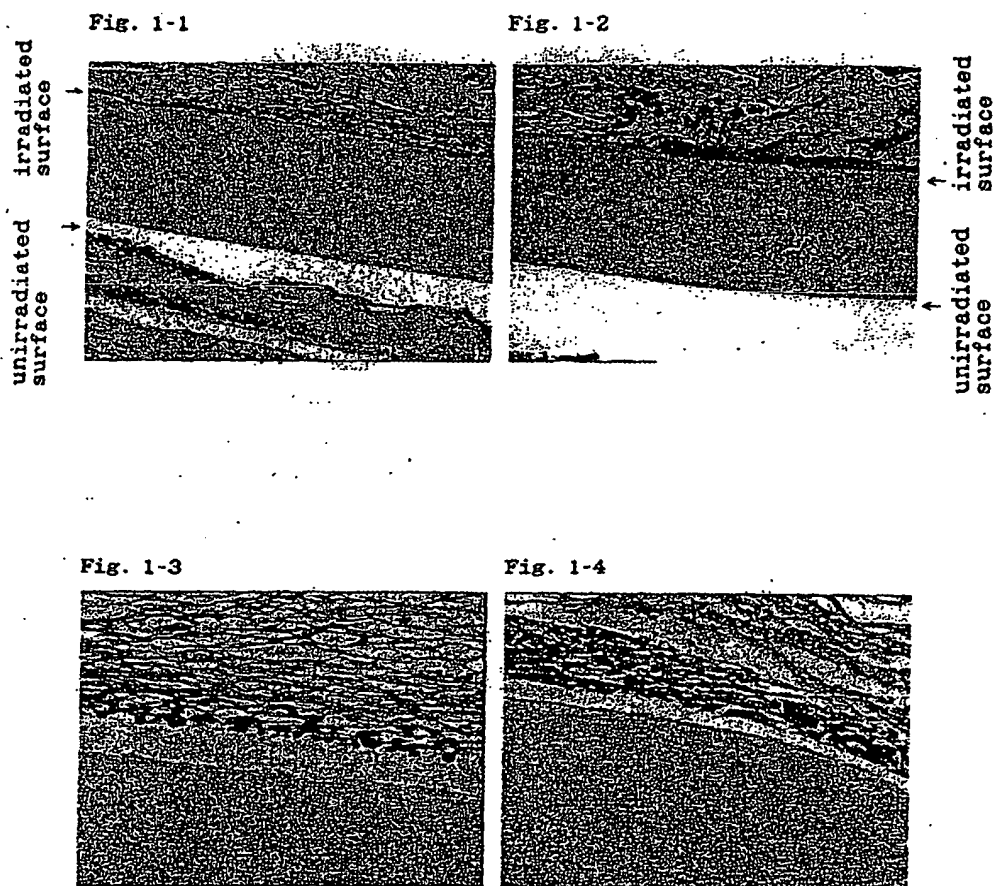
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[Fig. 1]



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[Fig. 2]

Fig. 2-1

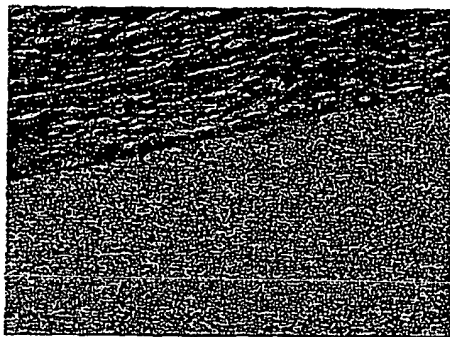


Fig. 2-2

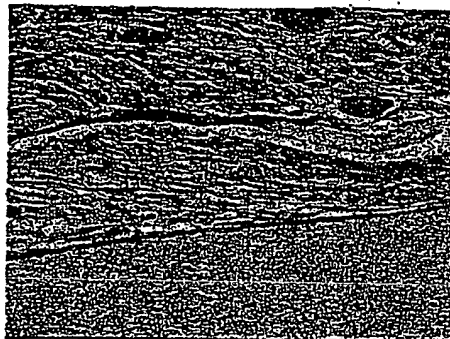
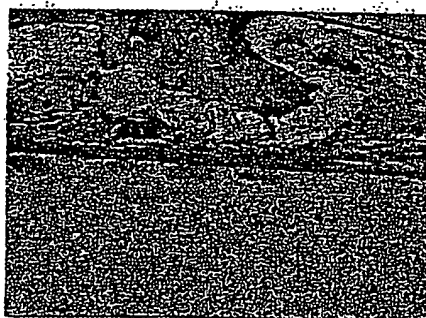


Fig. 2-3



Fig. 2-4



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[Fig. 3]

Fig. 3-1

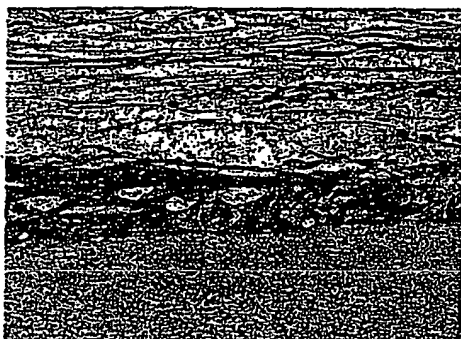


Fig. 3-2

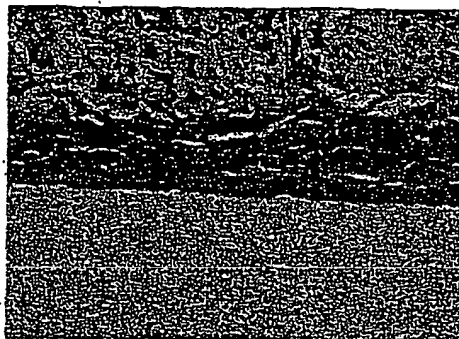


Fig. 3-3

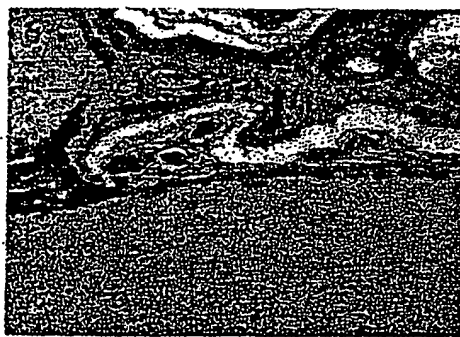


Fig. 3-4



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[Fig. 4]

Fig. 4-1



Fig. 4-2

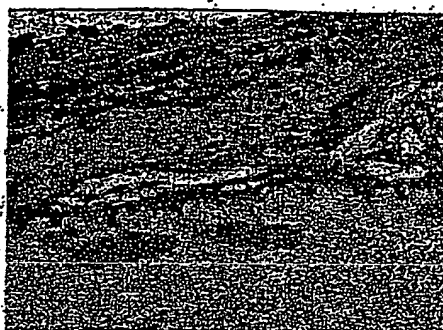


Fig. 4-3



Fig. 4-4

